## **DNA Precipitation and Hybridization (FISH)**

## Section of Cancer Genomics, Genetics Branch, NCI National Institutes of Health

## **Reagents**

**Dextran Sulfate (50%)** 

Intergen, Cat. S4030

Ethanol, absolute

Formamide, deionized

Ambion, Cat. 9342

HCl, 1N

Human Cot-1 TM DNA (1mg/ml)

GIBCO BRL, Cat. 15279-011, 500 μg

**Rubber Cement** 

**Salmon Testes DNA** (stock ~10 mg/ml)

SIGMA Molecular Biology, Cat. D-7657, 1ml

Sodium acetate, 3M (NaAcetate)

20X SSC

# **Preparation**

#### Master mix

Dextran sulfate, 50% 40 ml 20X SSC, pH 7 10 ml Sterile dH<sub>2</sub>O 50 ml

Vortex solution and place tube on a shaking platform overnight to insure proper mixing.

Aliquot and store at -20°C

#### 70% Formamide/2X SSC

10 ml 20 X SSC

20 ml dH<sub>2</sub>0

70 ml deionized formamide

Adjust to pH 7 with 1N HCL

Aliquot and store at -20°C.

### **Procedure: Precipitation**

1. Add to an eppendorf tube:

Probe DNA (600 ng -1µg DNA)

Human Cot-1 DNA (note 1) 10 μl

Salmon sperm DNA 1 µl

- 2. Add Na-Acetate, 1/10 of the total volume of probe DNA mixture (above).
- 3. Add 100% ethanol, i.e., 2.5 x total volume of mixture of DNA + NaAcetate.
- 4. Vortex, store tube at -20°C overnight or at -80°C for 30 min.
- 5. Centrifuge (13,000 rpm) the precipitated DNA at 4°C for 30 min.
- 6. Carefully pour off supernatant, and place in a speed vac for 10 min (medium heat) until pellet is dry.
- 7. Add 5 μl deionized formamide (pH 7.0) and incubate tube at 37°C using a thermomixer, shaking, for 30 min, and vortex several times during the incubation.
- 8. Add 5 μl Master Mix, vortex, centrifuge briefly.
- 9. Denature probe DNA at 80°C for 5 min, centrifuge briefly.
- 10. Pre-anneal probe at 37°C for 1 hr.

### **Procedure: Slide Denaturation and Hybridization**

- 1. Apply 120 μl 70% deionized formamide/2X SSC to a 24 mm x 60 mm coverslip. Touch the slide to the coverslip.
- 2. Denature slide at 80°C on a hot plate for 1.5 min (see note 1).
- 3. Quickly and carefully remove the coverslip and immediately place the slide in ice cold 70% ethanol, followed by 90% ethanol and 100% ethanol (for 3 min each).
- 4. Allow slides to air dry.

- Add pre-annealed probe DNA to the denatured slide and cover with an 18 mm<sup>2</sup> coverslip, carefully remove any bubbles with forceps, and completely seal the edges of the coverslip with rubber cement.
- 6 Hybridize at 37°C in hybridization chamber (use a light tight chamber if the probe is directly labeled) for 48 hr.

## Notes

- 1. cDNA probes do not require Cot-1 DNA.
- 2. The denaturation time depends on the age of the slide. For slides older then 30 days a denaturation time of 2 min is recommended.